

Influence of Elevated Carbon Dioxide on Interactions Between *Frankliniella occidentalis* and *Trifolium repens*

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ABSTRACT Elevated CO₂ concentrations can increase plant growth and change plant nutritive value for herbivores. Several reports indicate that leaf-chewing insects consume more foliage of plants grown at elevated CO₂ concentrations than of plants grown at ambient CO₂. Research with additional plant-pest systems is needed to determine if this phenomenon is widespread and if increased insect feeding might affect productivity. Effects of CO₂ enrichment on foliar consumption and population size of Western flower thrips [*Frankliniella occidentalis* (Pergande)] were measured on white clover (*Trifolium repens* L.). White clover infested with thrips was exposed for 24 h/d to ≈396 (ambient) or 745 μLL⁻¹ (elevated) CO₂ for up to 35 d in 10 greenhouse exposure chambers. At elevated CO₂, clover shoot weight and laminae weight were ≈50% greater, and laminar area was ≈20% greater than at ambient CO₂. Thrips population size was not significantly affected by CO₂, but laminar area scarred by thrips feeding was ≈90% greater at elevated than at ambient CO₂. Because of increased growth, however, undamaged leaf area was approximately 15% greater at elevated than at ambient CO₂.

KEY WORDS *Trifolium repens*, white clover, carbon dioxide enrichment, *Frankliniella occidentalis*, Western flower thrips

INCREASED ATMOSPHERIC CARBON DIOXIDE (CO₂) affects plant photosynthesis and chemistry (Rogers et al. 1983a, Cure and Aycock 1986, Kimball 1986), thereby influencing plant tissue nutritive quantity and quality for arthropods. Leaves of green plants at elevated CO₂ generally contain higher percentage soluble carbohydrates and lower percentage N than those at ambient CO₂ (Watt et al. 1995, Bezemer and Jones 1998, Coviella and Trumble 1999). Leaf chewing insects often consume more foliage of plants grown in CO₂-enriched air than of plants grown at ambient CO₂, possibly to compensate for decreased foliar N (Lincoln et al. 1986, Weste et al. 1987, Lincoln 1993, Watt et al. 1995, Brooks and Whittaker 1998, Buse et al. 1998, Lindroth and Kinney 1998, Stiling et al. 1999). The few studies of CO₂ enrichment effects on populations of whole-cell feeding arthropods have provided mixed results. Carbon dioxide enrichment suppressed populations of greenhouse whiteflies [*Trialeurodes vaporariorum* (Westward)] on tomato (Tripp et al. 1992) but increased populations of the twospotted spider mite (*Tetranychus urticae* Koch) on white clover (Heagle et al. 1994a, 2002). Populations of sweet potato whitefly [*Bemisia tabaci* (Gennadius)] on cotton (Butler et al. 1986) and of Western flower

thrips [*Frankliniella occidentalis* (Pergande)] on milkweed (*Asclepias syriaca* L.) (Hughes and Bazzaz 1997) were not affected by CO₂ enrichment. Thrips (probably *F. occidentalis*) populations in cotton *Gossypium hirsutum* L. also were not significantly affected by CO₂ enrichment of their cotton host (Butler 1985).

Western flower thrips (*F. occidentalis*) is one of the most important and difficult to control plant pests. It feeds on numerous plant species and spreads the tomato spotted wilt virus, which also affects numerous plant species. One experiment showed that consumption of *A. syriaca* leaves by *F. occidentalis* was significantly greater on plants exposed to elevated CO₂ compared with ambient levels, although the population size was not significantly affected (Hughes and Bazzaz 1997).

During recent greenhouse experiments, a severe infestation of *F. occidentalis* precluded further work to determine effects of mixtures of O₃ and CO₂ on white clover (*Trifolium repens* L.). This infestation provided an opportunity to study the effects of CO₂ enrichment on interactions between *F. occidentalis* and *T. repens*. Our objective was to determine if CO₂ enrichment affects foliar consumption and population size of *F. occidentalis* on *T. repens*.

Materials and Methods

General. The experiment was performed in a non-filtered-air greenhouse 5 km south of Raleigh, NC,

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Table 1. Carbon dioxide concentrations, temperature, relative humidity (RH), and PAR during exposures of white clover and thrips to carbon dioxide^a

| Trial | Dates of exposure | Carbon dioxide concentration (μLL^{-1}) | | Temperature ($^{\circ}\text{C}$) | RH (%) | PAR ($\text{mol m}^{-2}/\text{d}$) |
|-------|-------------------|------------------------------------------------------|----------------|------------------------------------|--------|--------------------------------------|
| | | Ambient | Double ambient | | | |
| 1 | 27 Feb.–26 Mar | 393 | 729 | 20 | 58 | 34 |
| 2 | 3 Apr.–7 May | 398 | 761 | 22 | 62 | 30 |

^a All values are 24-h means. Values for CO_2 concentrations across replicate chambers are within 3% of values shown. Values for temperature across replicate chambers were within 1°C of values shown. Values for RH across replicate chambers were within 1% of values shown. Values for PAR across chambers were within 9% of values shown.

between late February and early May. A *T. repens* clone (NC-S) previously described (Heagle et al. 1993, 1994b) was propagated by rooting virus-free stolons in 1-liter pots containing Metro Mix 220 and 4 g of slow release Osmocote fertilizer (14–14–14, N-P-K; Scotts-Sierra Horticultural Products, Marysville, OH). Plants were inoculated with commercial rhizobium inoculant 14 d after start of rooting and were watered as needed to prevent wilting throughout the experiment.

Exposures. Plants were exposed to ambient and approximately double ambient CO_2 concentrations in 10 cylindrical (1.07 m diameter by 1.20 m tall) continuous-stirred tank reactor chambers (CSTRs) continuously aspirated with charcoal-filtered air (Heck et al. 1978). Ambient greenhouse light was supplemented by 1,000-W multivapor lamps that supplied $\approx 270 \mu\text{mol}/\text{m}^2/\text{s}$ of photosynthetically active radiation (PAR) to each chamber from 0600 to 1800 h EST daily. Temperature was monitored in the CSTRs with copper-constantan thermocouples, PAR was measured with quantum sensors (LI-190S-1; LI-COR, Lincoln, NE), and relative humidity was measured with Vaisala HMP 31UT sensors (Vaisala, Woburn, MA).

A naturally occurring population of *F. occidentalis* was present on *T. repens* growing in 15-liter pots. New plants were rooted as described above in 1-liter pots on a bench between two benches that each contained ten 15-liter pots of the thrips-infested plants. Evidence of thrips injury on new plants appeared as irregular white necrotic areas and distortion of newly expanding trifoliolates within 3 wk after rooting began.

The experimental design was a randomized complete block with five blocks of the two CO_2 concentrations as the main plot (CSTR) treatments with nine 1-liter pots of clover randomly assigned to each of the 10 CSTRs. Plants were moved to the CSTRs 26 d after rooting began for trial 1 and 21 d after rooting began for trial 2. Exposures to CO_2 for 24 h/d began 1 d after plants were moved to the CSTRs. CO_2 was obtained from tank CO_2 and was monitored sequentially in each chamber with an infrared CO_2 analyzer (LI 6252 (LI-COR). Details of CO_2 dispensing and monitoring protocols used in this study have been described (Rogers et al. 1983b). Exposures continued for 27 d for trial 1 and for 35 d for trial 2. Mean concentrations of CO_2 , temperature, PAR, and relative humidity during CO_2 exposure for each trial are shown in Table 1.

Measurements. Six plants in each CSTR were used to measure thrips populations, and three plants in each

chamber were used to measure area and weight of leaf laminae and to estimate foliar scarring by thrips. To measure thrips populations, we modified a procedure for collecting thrips on fresh pole bean pods previously described (Groves et al. 2001). One day before exposures ended, foliage and stems from three two-plant samples per chamber were cut and placed in white plastic containers (20 cm diameter by 16 cm tall). To ensure air exchange, organically cloth was used to replace a 10-cm-diameter area of the bottom and the entire top of each container. One fresh pole bean pod (*Phaseolus vulgaris* L.) was placed at the bottom of each container as a food and moisture source for the thrips. The containers were placed on their sides in a desiccation chamber set at 28°C and 20% RH. For trial 1, the clover foliage was dry after 3 d. Thrips still among the dried foliage in each container were encouraged to move downward toward the bean pod by exhaling into the container while gently rustling and slowly removing the dried foliage. Bean pods with thrips were placed in ethyl alcohol (96%), and adults and immature thrips were counted later using a microscope. The dried foliage for each two-plant sample was weighed. For trial 2, the procedure was similar except drying required 7 d because plants were larger than for trial 1. For trial 2, plant material from each container (<1 g) remaining after bean pods were collected was placed in plastic bags. Adults and immature thrips in this sample and in ETOH were counted with a microscope.

Within 3 h after exposures ended, area of leaf laminae was measured for three plants per chamber using an electronic meter (LI-3100; LI-COR). Thrips feeding on expanded leaves caused irregular pale scarred areas on abaxial leaf surfaces. The percentage scarred area was visually estimated for each leaflet (0–100%). Total leaf area scarred per plant was calculated as mean area per leaflet \times mean percentage area per leaflet scarred by thrips \times number of leaflets per plant. Leaf laminae were dried at 50°C and weighed.

Statistical Analyses. Analyses of variance (ANOVA) were performed on chamber means. Analysis of the trials separately showed similar responses to treatments. Data from both trials were then combined and analyzed with trial considered to be a fixed factor. Residual plots were examined for non-normality, outliers, and heterogeneous variances. The Box-Cox test indicated that percentage leaf area scarred by thrips was best analyzed using the log transformation. All other variables were analyzed using original scales.

Table 2. Effects of CO₂ enrichment on *T. repens* growth and foliar consumption of *T. repens* by *F. occidentalis*^a

| Trial | CO ₂ concentration μLL ⁻¹ | Shoot dry weight per 2-plant sample (SE) ^a (g) | Laminae wt per plant (SE) (g) | Number of leaflets per plant (SE) | Laminar area per plant (SE) (cm ²) | Thrips per 2-plant sample ^a | | Laminar area scarred per plant ^b (cm ²) |
|-------------------------|----------------------------------------------------|-----------------------------------------------------------------|-------------------------------------|-----------------------------------------|------------------------------------------------------|-------------------------------------------|----------------|-------------------------------------------------------------------------|
| | | | | | | immatures (SE) | adults (SE) | |
| 1 | 393 | 13.5 (0.14) | 4.6 (0.11) | 127 (1.6) | 628 (17.7) | 278 (27.9) | 21 (1.7) | 31 (3.6) |
| | 729 | 20.1 (1.39) | 6.8 (0.23) | 127 (1.0) | 748 (25.9) | 270 (36.7) | 13 (0.9) | 60 (4.4) |
| 2 | 398 | 17.6 (0.33) | 6.5 (0.23) | 170 (2.0) | 1017 (39.6) | 533 (060.2) | 27 (3.0) | 73 (15.0) |
| | 761 | 27.1 (1.02) | 10.3 (0.28) | 187 (1.2) | 1232 (19.1) | 516 (143.5) | 32 (7.6) | 139 (29.6) |
| Source | df | | | | | | | |
| | | | | Probability of >F value from ANOVA | | | | |
| Trial | 1 | 0.0003 | 0.0001 | 0.0001 | 0.0001 | 0.0249 | 0.0092 | 0.0430 |
| CO ₂ | 1 | 0.0001 | 0.0001 | 0.0005 | 0.0001 | 0.8630 | 0.7854 | 0.0007 |
| Trial × CO ₂ | 1 | 0.1326 | 0.0111 | 0.0007 | 0.1165 | 0.9512 | 0.2092 | 0.8919 |

^a Each value is the mean of 15 samples (three two-plant samples or three one-plant samples in each of five replicate chambers).

^b Laminar area scarred per plant calculated as (mean area per leaflet × mean percentage area scarred per leaflet × number of leaflets).

Results

Plants were larger in trial 2 than trial 1, and the trial effect was significant for all responses measured (Table 2). Plants at elevated CO₂ were larger than at ambient CO₂, and the CO₂ effect was significant for all plant responses measured. Shoot weight and leaf lamina weight were ≈50% greater in elevated than at ambient CO₂ in both trials. Leaf numbers were not affected by CO₂ in trial 1 but were greater at elevated than at ambient CO₂ in trial 2, accounting for the significant trial × CO₂ effect. Laminar area was ≈20% greater at elevated than at ambient CO₂ in both trials.

Thrips populations were ≈1.3 times greater in trial 2 than in trial 1, but population size was not significantly affected by CO₂ in either trial (Table 2). However, laminar area per plant scarred by thrips feeding was ≈90% greater in elevated than ambient CO₂ in both trials (Table 2). In trial 1, per capita leaf area scarred (laminar area scarred per plant divided by total thrips population per plant) averaged 0.21 cm² at ambient CO₂ and 0.42 cm² at elevated CO₂. Comparable values for trial 2 were 0.26 and 0.51 cm², respectively.

Discussion

The current study essentially corroborates previous results showing increased per capita feeding of milkweed by Western flower thrips with no significant thrips population change (Hughes and Bazzaz 1997). Neither the present or previous results should be considered proof that elevated CO₂ does not affect thrips populations, however. A major reason is that neither study simulated seasonal exposure duration. Exposures of clover lasted 27 d in trial 1 and 35 d in trial 2 at temperatures averaging 20–22°C. Exposure of milkweed was ≈28 d (Hughes and Bazzaz 1997). These durations allowed time for only an estimated 1.5–2.0 generations (Martin 1993, Baker 1994).

Nutritive changes may be partly responsible for effects of CO₂ enrichment on herbivores, but evidence is mostly limited to correlations between insect response and whole-leaf analyses showing increased C, C/N ratios, and carbohydrates accompanied by

decreased N. Foliar nutritive levels were not measured in the present experiment. However, previous analyses of leaves of *T. repens* (NC-S) showed that elevated CO₂ decreased percentage N by 13%, increased nonstructural carbohydrates by 49%, and did not significantly affect the concentration of any of 14 amino acids (Heagle et al. 2002).

Most research measuring effects of elevated CO₂ has included only two CO₂ concentrations (ambient and double ambient). It is not possible to accurately estimate effects of intermediate CO₂ concentrations from studies using only two concentrations because responses to CO₂ may or may not be linear. Because ambient CO₂ concentrations have been rising over the past century, the two-concentration approach does little to show if increases in atmospheric CO₂ that have already occurred are causing significant effects on plants or plant pests. Dose-response designs with multiple CO₂ concentrations are needed to develop models that can estimate effects at all CO₂ concentrations.

Whereas consumption of white clover by thrips at elevated CO₂ was ≈90% greater than that at ambient CO₂, total leaf area increased by ≈20%, and undamaged leaf area increased by ≈15%. This net increase in undamaged leaf area occurred despite feeding by a relatively high thrips population. Whether or not elevated CO₂ will be a net benefit for *T. repens* under field conditions remains to be determined.

The present results show that increased feeding by *F. occidentalis* in response to CO₂ enrichment is not limited to a given plant species. Because both *F. occidentalis* and tomato spotted wilt virus have an extremely wide host range, the possibility that elevated CO₂ might increase the spread and prevalence of tomato spotted wilt virus should be investigated.

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References Cited

- Baker, J. R. 1994. Western flower thrips. Ornamentals and turf insect Note 72 (EnT/ort-72). North Carolina Cooperative Extension Service, North Carolina State University, Raleigh, NC.
- Bezemer, T. M., and T. H. Jones. 1998. Plant-insect herbivore interactions in elevated atmospheric CO₂: quantitative analyses and guild effects. *Oikos* 82: 212–222.
- Brooks, G. L., and J. B. Whittaker. 1998. Responses of multiple generations of *Gastrophysa viridula*, feeding on *Rumex obtusifolius*, to elevated CO₂. *Global Change Biol.* 4: 63–75.
- Buse, A., J.E.G. Good, S. Dury, and C. M. Perrins. 1998. Effects of elevated temperature and carbon dioxide on the nutritional quality of leaves of oak (*Quercus robur* L.) as food for the winter moth (*Operophtera brumata* L.). *Funct. Ecol.* 12: 742–749.
- Butler, G. D., Jr. 1985. Populations of several insects on cotton in open-top carbon dioxide experiment chambers. *Southwestern Entomologist*. 10: 264–267.
- Butler, G. D., Jr., B. A. Kimball, and J. R. Mauney. 1986. Populations of *Bemisia tabaci* (Homoptera:Aleyrodidae) on cotton grown in open-top field chambers enriched with CO₂. *Environ. Entomol.* 15: 61–63.
- Cure, J. D., and B. Aycok. 1986. Crop responses to carbon dioxide doubling: a literature survey. *Agric. Forest. Meteorol.* 38: 127–145.
- Coviella, C. E., and J. T. Trumble. 1999. Effects of elevated atmospheric carbon dioxide on insect-plant interactions. *Conserv. Biol.* 13: 700–712.
- Groves, R. L., C. E. Sorenson, J. F. Walgenbach, and G. G. Kennedy. 2001. Effects of imidacloprid on transmission of tomato spotted wilt tospovirus to pepper, tomato and tobacco by *Frankliniella fusca* Hinds (Thysanoptera: Thripidae). *Crop Protect.* 20: 439–445.
- Heagle, A. S., R. L. Brandenburg, J. C. Burns, and J. E. Miller. 1994a. Ozone and carbon dioxide effects on spider mites in white clover and peanut. *J. Environ. Qual.* 23: 1168–1176.
- Heagle, A. S., J. C. Burns, D. S. Fisher, and J. E. Miller. 2002. Effects of carbon dioxide enrichment on leaf chemistry and reproduction by twospotted spider mites on white clover. *Environ. Entomol.* 31: 594–601.
- Heagle, A. S., J. E. Miller, and D. E. Sherrill. 1994b. A white clover system to estimate effects of tropospheric ozone on plants. *J. Environ. Qual.* 23: 613–621.
- Heagle, A. S., J. E. Miller, D. E. Sherrill, and J. O. Rawlings. 1993. Effects of ozone and carbon dioxide mixtures on two clones of white clover. *New Phytologist*. 123: 751–762.
- Heck, W. W., R. B. Philbeck, and J. A. Dunning. 1978. A continuous stirred tank reactor (CSTR) system for exposing plants to gaseous air contaminants: Agricultural Research Service, U. S. Dep. Agriculture, ARS-S-181, New Orleans, LA.
- Hughes, L., and F. A. Bazzaz. 1997. Effect of elevated CO₂ on interactions between the western flower thrips, *Frankliniella occidentalis* (Thysanoptera: Thripidae) and the common milkweed, *Asclepias syriaca*. *Oecologia* (Berl.). 109: 286–290.
- Kimball, B. A. 1986. CO₂ stimulation of growth and yield under environmental constraints, pp. 53–57. In H. Z. Enoch and B. A. Kimball (ed.), Carbon dioxide enrichment of greenhouse crops, vol. II. Physiology, yield and economics. CRC. Boca Raton, FL.
- Lincoln, D. E. 1993. The influence of plant carbon dioxide and nutrient supply on susceptibility to insect herbivores. *Vegetatio* 104/105: 273–280.
- Lincoln, D. E., D. Couvet, and N. Sionit. 1986. Response of an insect herbivore to host plants grown in carbon dioxide enriched atmospheres. *Oecologia* (Berl.). 69: 556–560.
- Lindroth, R. L., and K. K. Kinney. 1998. Consequences of enriched atmospheric CO₂ and defoliation for foliar chemistry and gypsy moth performance. *J. Chem. Ecol.* 24: 1677–1695.
- Martin, N. A. 1993. Western flower thrips 1. Biology, identification and life cycle. Crop & Food Research Broad-sheet 35. The New Zealand Institute for Crop & Food Research Limited, Christchurch, NZ.
- Rogers, H. H., G. E. Bingham, J. D. Cure, J. M. Smith, and K. A. Surano. 1983a. Responses of selected plant species to elevated carbon dioxide in the field. *J. Environ. Qual.* 12: 569–574.
- Rogers, H. H., W. W. Heck, and A. S. Heagle. 1983b. A field technique for the study of plant responses to elevated carbon dioxide concentrations. *J. Air Pollut. Control Assoc.* 33: 42–44.
- Stiling, P., A. M. Rossi, B. Hungate, P. Dijkstra, C. R. Hinkle, W. M. Knott, and B. Drake. 1999. Decreased leaf-miner abundance in elevated CO₂: reduced leaf quality and increased parasitoid attack. *Ecolog. Applic.* 9: 240–244.
- Tripp, K. E., W. K. Kroen, M. M. Peet, and D. H. Willits. 1992. Fewer whiteflies found on CO₂ enriched greenhouse tomatoes with high C:N ratios. *Hort. Sci.* 27: 1079–1080.
- Watt, A. D., J. B. Whittaker, M. Docherty, G. Brooks, E. Lindsay, and D. T. Salt. 1995. The impact of elevated atmospheric CO₂ on insect herbivores, pp. 197–219. In R. Harrington and N. E. Stork (eds.), Insects in a changing environment. Academic, London.
- Weste, L., A. Osbrink, J. T. Trumble, and R. E. Wagner. 1987. Host suitability of *Phaseolus lunata* for *Trichoplusia ni* (Lepidoptera: Noctuidae) in controlled carbon dioxide atmospheres. *Environ. Entomol.* 16: 639–644.

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